
Mammary gland development and cancer

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Summary

The mammary gland is a complex organ that begins development early in gestation and constantly changes in size, shape and function from the time of puberty to menopause. The earliest stages of embryogenesis appear to be independent of steroid hormones, whereas after the 15th week breast structure is largely influenced by a variety of hormones. In most females, further breast development begins at puberty under the influence of cyclical estrogen and progesterone secretion. This process may continue into the 20s and it is enhanced by pregnancy. Growth and transcription factors contribute to the reciprocal stromal-epithelial interactions in growth, development and tumorigenesis of the mammary gland. From the embryological point of view the morphology of both mammary ductal and lobular cells results from the same developmental process. Numerous data suggest the existence of self-renewing, pluripotent mammary stem cells but their molecular characteristics and differentiation pathways are unknown. The extensive research currently being done in molecular biology and pathology, cancer genomics and proteomics will hopefully contribute to further elucidation of all the genetic and environmental factors involved in the development, differentiation, and involution of the mammary gland and this may give insight into the etiopathogenesis, early detection, treatment, and potential prevention of breast cancer.

Key words: mammary gland development – differentiation – growth factors – transcription factors – ductal and lobular cells – breast cancer

Souhrn

Prsní žláza – vývoj a nádory

Prsní žláza je komplexní orgán, jehož vývoj začíná záhy v gestaci, a který se od puberty k menopauze průběžně mění ve velikosti, tvaru i funkci. Zatímco nejčasnější stadia embryogeneze probíhají nezávisle na steroidních hormonech, po 15. týdnu je struktura prsu významně ovlivněna řadou hormonů. U většiny žen začíná po pubertě další vývoj prsu pod vlivem cyklické sekrece estrogenu a progesteronu. Tento proces může pokračovat i po dvacítce a je potencován těhotenstvím. Růstové a transkripční faktory přispívají ke vzájemným interakcím stroma – epitel při růstu vývoji normální prsní žlázy, i při patogenezi. Z pohledu embryologie vychází morfologie duktálních i lobulárních buněk prsu z téhož vývojového procesu. Existují četné údaje o existenci obnovujících se pluripotentních kmenových buněk prsní žlázy, avšak jejich molekulární charakteristika a cesty diferenciace nejsou dosud známy. Lze doufat, že probíhající rozsáhlý výzkum molekulární biologie, patologie a nádorové genomiky a proteomiky přispěje k objasnění všech genetických i zevních faktorů účastnících se vývoje, diferenciace a involuce prsní žlázy, a zároveň povede k lepšímu pochopení etiopatogeneze, časnému záchytu, léčbě a možná i prevenci karcinomu prsu.

Klíčová slova: prsní žláza – vývoj – diferenciace – růstové faktory – transkripční faktory – duktální a lobulární buňky – karcinom prsu

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Mammary gland embryogenesis

Over the past few years, significant new insights have been made in our understanding of the transcription factors that guide development

of the mammary gland. The human mammary glands develop from the mammary ridges which first appear in 5-week fetus. The ridge does not develop further and disappears during fetal development (22, 54). On the chest wall, mesenchymal condensation occurs around an epithelial stalk at about the 15th week. Further

development of the buds into ductal structures involves reciprocal signaling between the epithelium, the mammary mesenchyme and the stromal fat pad (56). Inhibins and activins regulate mammary epithelial cell differentiation through mesenchymal-epithelial interactions (53). Growth of cords of epithelium into the mesenchyme produces a group of solid epithelial columns, each of which give rise to a lobe in the mammary gland. The papillary layer of the fetal dermis gives rise to vascularized fibrous tissue (40). Coincidentally, differentiation of the mesenchyme into fat within the last 2 months of gestation and canalization of the epithelial cords occur, followed by development of branching lobuloalveolar glandular structures. Near birth, the nipple is formed. Between birth and puberty the mammary gland is relatively quiescent, with only limited ductal elongation into the mammary fat pad (22, 23).

LeBrun et al. found that the expression of bcl-2 protein is more widespread in fetal than adult tissues, which is consistent with its role in promoting cell survival, presumably by preventing apoptosis (16, 33, 38). Immunohistochemical localization of Bcl-2 has been detected in the basal epithelium of the developing breast bud and in the surrounding stroma. Similar patterns of staining have been reported in male and female tissue. Bcl-2 is not present in the epithelium of the normal adult breast. These observations suggest that up-regulation of bcl-2 contributes to morphogenesis of the fetal breast by its inhibitory effect on apoptosis (44).

Postnatal development and hormonal regulation of mammary gland

The earliest stages of fetal mammary gland development appear not to be dependent on steroid hormones, whereas the actual development of the breast structure after the 15th week is largely influenced by testosterone (22). In the last weeks of gestation, the fetal breast is responsive to steroid hormones and prolactin, which is manifested after birth by the secretion of colostrum and palpable enlargement of the breast bud. The secretory activity typically subsides and ceases during the first or second month after birth (23, 26).

The morphological changes that the mammary gland undergoes during puberty, pregnancy and lactation are primarily defined by steroid and peptide hormones (42). In most females, further breast development does not begin until puberty (70). With the onset of cyclical estrogen and progesterone secretion at puberty,

adolescent breast development commences and involves rapid ductal outgrowth and branching (53). Branching emanates from the terminal end buds, which are specialised regions occurring at the tips of the growing ducts. These contain the precursor cells to the luminal epithelial and myoepithelial cells. Growth of ducts and differentiation of periductal stroma occur at this time in estrogen-dependent manner (2, 69). Growth hormone and glucocorticoids contribute to ductal growth. Lobuloalveolar differentiation and growth during this period are enhanced primarily by insulin, progesterone, and growth hormone. By the end of puberty, a simple ductal system or primary and secondary ducts are developed (58, 59).

Anbazhagan et al. devised a system of classification to study the extent of the structural development of the ductal system (morphological types I, II, and III) and the functional differentiation of the lining epithelium (functional stages I to V) in infant breasts collected at necropsy. There was no correlation between the age of the infant and the type of development of the ductal system. There were no distinguishing features between breasts from the two sexes. Myoepithelial cells were present at all stages and prominent staining for casein was observed up to 2 months of age (1).

Breast glandular differentiation may continue into the 20s and it is enhanced by pregnancy. During pregnancy, further ductal side-branching occurs together with lobuloalveolar proliferation leading to the formation of the lobuloalveolar compartment with secretory epithelium. This phase of development is driven by progesterone, prolactin and other pregnancy-induced hormones (43, 45). Stat5 (signal transducer and activator of transcription) pathway is critical for the determination, proliferation, and differentiation of mammary alveoli during pregnancy (25). Geymayer and Doppler provide evidence for a role of NF-kappaB (nuclear factor kappa-B) in normal mammary gland development, and indicate its function as a negative regulator of beta-casein gene expression during pregnancy by interfering with STAT5 tyrosine phosphorylation (19). Miyoshi et al. demonstrated that signalling via the PrlR and Stat5 is critical for the proliferation and differentiation of mammary alveoli during pregnancy (41). The observations of Philp et al. revealed a complex pattern of activation of STAT factors during mammary growth, differentiation and remodelling and provided the evidence for the involvement of STAT3 in development of the mammary gland (49). It has been shown that during pregnancy beta-casein synthesis is induced in single mammary epithelial cells via an integrin-dependent pathway (27, 53). Beta1-

integrins play an important role in the control of milk gene transcription and in the maintenance of the mammary epithelial cell differentiated state (15).

SRC-1 (steroid receptor coactivator-1) family members interact with steroid receptors, including ER α (estrogen receptor alpha) and PR (progesterone receptor), to enhance ligand-dependent transcription. Shim et al. documented that SRC-1 is not necessary for ER α -mediated induction of PR in mammary epithelial cells and is also not sufficient for PR induction in stromal cells expressing both ER α and SRC-1 (63). Chughtai et al. report that Src homology 2 (SH2) domain containing protein-tyrosine phosphatase SHP-2 contributes to prolactin receptor (PRLR) signal transduction to beta-casein gene promoter activation. These studies indicate a tight physical and functional interaction between SHP2 and Stat5 required for PRL-mediated signalling in mammary cells (7). It has been shown that p21-activated kinase (Pak) 1, a serine/threonine protein kinase, is activated in mammary glands during pregnancy and lactation (72). Following weaning, the mammary gland undergoes extensive remodeling through apoptosis of the secretory cells, returning the gland to a virgin-like state (22, 59).

Saji et al. found that ER α and ER β are expressed in the rat mammary gland but the presence and cellular distribution of the two receptors are distinct. In prepubertal rats, ER α was detected in 40 % of the epithelial cell nuclei. This decreased to 30 % at puberty and continued to decrease throughout pregnancy to a low of 5 % at day 14. Cells coexpressing ER α and ER β were rare during pregnancy, a proliferative phase, but they represented up to 60% of the epithelial cells during lactation, a postproliferative phase (60). Forster et al. suggest a role for ER β in organization and adhesion of epithelial cells and hence for differentiated tissue morphology. Because a reduced risk for breast cancer is conferred on women who breast-feed at an early age, ER β could contribute to this risk reduction by facilitating terminal differentiation of the mammary gland (18). Russo et al. concluded that the content of ER α and PR in the normal mammary tissue varies with the degree of lobular development, in parallel with cell proliferation. However, the expression of receptors occurs in cells other than the proliferating cells, indicating that they represent at least two separate cell populations (57). Shyamala et al. found that PR present in a heterogeneous manner in breast epithelial cells are essential for lobuloalveolar and not for ductal morphogenesis. Progesterone signalling through PR may occur through paracrine mechanism and regulated expression of the two isoforms of PR is critical for

maintaining appropriate responsiveness to progesterone and epithelial cell replicative homeostasis (64).

Growth factors in mammary development and carcinogenesis

Growth factors contribute to the reciprocal stromal-epithelial interactions in growth, development and tumorigenesis of the mammary gland. Signalling by members of the epidermal growth factor receptor family plays an important role in breast development and breast cancer. It has been shown that EGFR (epidermal growth factor receptor) is essential for mammary ductal growth and branching morphogenesis, but not for mammary lobulo-alveolar development (73, 74). Cripto-1 (CR-1) also known as teratocarcinoma-derived growth factor-1 is a novel EGF-related protein that induces branching morphogenesis in mammary epithelial cells and inhibits the expression of various milk proteins. CR-1 is also overexpressed in mammary tumors (62).

The ERBB (receptor protein-tyrosine kinase) family of type 1 receptor tyrosine kinases and their ligands play important role during mammary morphogenesis. ERBB4 functions as an essential mediator of the mammary differentiation factor STAT5 signalling, and that loss of STAT5 activity contributes to the impaired functional differentiation of mammary glands observed in mice containing conditional *ErbB4* deletions (37). Jones et al. found that *ErbB4* signalling is necessary for terminal differentiation of mouse mammary gland and Stat5 activation at mid-lactation (28). Kazansky et al. detected immunocytochemically considerable cytoplasmic and some nuclear staining for Stat5a1 during late pregnancy and predominantly nuclear staining during early lactation (29). Yamashita et al. proposed that serine phosphorylation within the transactivation domain may limit the activity of Stat5a in the absence of proper coactivation by glucocorticoid receptors. Costimulation of glucocorticoid receptors completely reverses the suppressive effect of Stat5a serine phosphorylation on beta-casein gene transcription (76).

Osin et al. found that the distribution of growth factors and extracellular matrix proteins play a significant role in different cellular compartments during morphogenesis of the developing human breast. TGF- α (transforming growth factor) immunoreactivity was observed both in the stromal and the epithelial cells within fetal and infant breasts up to 25 days. TGF- β 1 immunoreactivity was localized in the extracellular matrix (46). In addition, tenascin-C

was found around the neck of the developing breast bud and in the extracellular matrix of the infant with peaks in the newborn at 6–12 weeks (61).

Keratinocyte growth factor/fibroblast growth factor 7 (KGF/FGF7) is known to be a potent growth factor for mammary cells but its origin, cellular targets and mode of action in the breast are unclear. Palmieri et al. showed epithelial and myoepithelial immunohistochemical staining for FGF7 in normal breast sections, and epithelial staining in breast carcinomas. FGF7 stimulated proliferation of both epithelial cell types, but not stromal fibroblasts. Interleukin-1 β caused a dose-related FGF7 release (47).

It has been shown that TGF- β 1 is located in the mammary gland periductal and intraductal stroma, closely associated with epithelial or myoepithelial cells in both benign and malignant breasts (3). The relative localization of these two growth factors in the mammary gland may be significant in the control of breast development and/or tumor formation and progression (20).

Recent studies have documented that PTHrP (parathyroid hormone-related protein) is a native product of mammary epithelial and myoepithelial cells and is necessary for mammary development (17). In the absence of a functional PTHrP gene, mammary development fails during embryonic life, and no mammary epithelial ducts are formed. This molecule participates in the regulation of epithelial-mesenchymal interactions not only during embryonic mammary development, but also during adolescent ductal morphogenesis. In addition, PTHrP plays a critical role in the establishment of bone metastases in breast cancer (13). The recent experiments have demonstrated that PTHrP and the PTH/PTHrP receptor represent an epithelial/mesenchymal signalling circuit that is necessary for mammary morphogenesis and stromal cells are a critical target for PTHrP's action in the mammary gland (14, 75).

Transcription factors in mammary gland development

The Ets (expressed sequence tags) gene family consists of 27 genes that encode sequence-specific transcription factors. Genetic analyses suggest roles for Ets proteins in embryonic development and various adult physiological processes (39). The overexpression of some ETs genes is linked with numerous malignancies, including breast cancer. The PEA3 (transcription factor polyomavirus enhancer activator protein)

subfamily proteins play key regulatory roles in both mammary gland development and oncogenesis (32). The bHLH (basic helix-loop-helix) family of transcription factors functions in the coordinated regulation of gene expression, cell lineage commitment, and cell differentiation in most mammalian tissues. Id proteins function as dominant negative regulators of bHLH transcription factors (9). Lin et al. identified Id-1 protein is a critical regulator of these normal mammary epithelial cell phenotypes, also regulator of aggressive and invasive phenotype and mediator of the effects of sex steroid hormones, in human breast cancer cells (36).

C/EBP α and - δ , the members of C/EBP (CCAAT/enhancer binding protein) family of bZIP transcription factors, are also expressed in mammary gland. The multiple protein isoforms of C/EBP β play a critical role in mammary gland development and cancer. Targeted deletion of all the C/EBP β isoforms results in a severe inhibition of lobuloalveolar development and a block to functional differentiation and ductal morphogenesis (21). Qi et al. found that nuclear receptor coactivator peroxisome proliferator-activated receptor-interacting protein (PRIP) is important for normal mammary gland development (50).

Mammary gland development is characterized by dynamic changes in the expression and functions of protein kinases. Perturbations in the regulated expression or function of protein kinases or their associated signalling pathways can lead to malignant transformation of the breast. Since receptor-tyrosine kinases regulate several essential processes such as mitogenesis, motility, invasion, cell survival, and angiogenesis, targeting receptor-tyrosine kinases may have important implications in designing strategies against breast cancer (31). Recent genetic and expression analyses strongly suggest that homeobox genes may perform similar functions at specific developmental transition points in the mammary gland. These analyses also suggest that homeobox genes may play a contributory or causal role in breast cancer (34).

To date, six main pathways affecting mammary morphogenesis have been described. These include: (1) LEF1 (lymphocyte enhancer factor 1) and b-catenin signalling, (2) homeobox genes *Msx1*, *Msx2* and *Hox* signalling (5), (3) steroid hormone receptor signalling, (4) STAT signalling, (5) Helix-loop-helix (HLH) gene signalling and (6) c-myc, Ets and LMO4 oncogene signalling (71). All can also be involved in breast cancer development (fig.1).

The data above discussed show that from the embryological point of view there exists no substantial difference between mammary ductal and lobular cells and that the morphology of both

results from the same developmental process. There is no evidence for a different gene expressing profile in normal ductal and lobular cells. However, some investigations report differential immunohistochemical markers of ductal and lobular cells.

Differential immunohistochemical markers of ductal and lobular cells

An essential feature of mammary gland biology is its ability to regenerate a functional epithelium upon successive cycles of lactation and involution. Determination of the cellular pathways through which this recapitulation of organ growth and differentiation occurs will provide an important framework for mammary gland conceptualizing mammary dysplasia and carcinogenesis (67). Numerous data suggest the existence of self-renewing, pluripotent mammary stem cells (8, 10); however, their molecular characteristics and differentiation pathways are largely unknown (30, 55). Transformed mammary stem or progenitor cells undergo aberrant differentiation processes that result in generation of the phenotypic heterogeneity found in human and rodent breast cancers (12).

It is suggested that epithelial/intermediate stem cells exist in a basal position predominantly in terminal structures of growing breasts. The acquisition of the malignant phenotype is associated with the carcinoma cells having a greatly impaired ability to differentiate to myoepithelial and to alveolar cells (55). Pechoux

et al. and Deugnier et al. suppose that human mammary luminal epithelial cells contain progenitors to myoepithelial cells (48, 11). Stingl et al. reported the evidence for the existence of multilineage HBEC (normal mammary epithelial cell line) progenitors in normal adult human mammary tissue. The MUC-1+/CALLA-/ESA+ and the MUC-1- to +/-CALLA +/- to +/ESA+ progenitors are candidates in vivo alveolar and ductal progenitors, respectively (68).

Recent transgenic mouse models have demonstrated that distinct pre-committed mammary epithelial progenitors function to regenerate mammary ducts and secretory mammary lobules. Lobular, i.e. secretory progenitor cells are present as entities among the mammary epithelial cells found in immature virgin female mice. Similarly, ductal epithelial progenitors are present within the same population. Lobular progenitors are present in greater numbers, although both cell populations are extremely small. The results may indicate cooperative interaction between the two epithelial progenitors or signal the presence of a third progenitor type, i.e. a mammary epithelial stem cell capable of producing both ductal and lobular-committed daughters (6, 35).

The study of Hebbard et al. suggests that the CD44v6 epitope is expressed in mammary epithelial stem cells and in lineages derived from these cells, and that CD44v6 expression is regulated in part by hormones and growth factors such as IGF-1 (insulin-like growth factor 1) and EGF (epidermal growth factor) which regulate the growth and differentiation of the mammary epithelium. The expression of the CD44v6 epitope observed in some mammary tumors reflects the stem cell origin of breast tumors (24).

14-3-3sigma is a candidate tumour suppressor gene trans-activated by p53 in response to DNA damage. 14-3-3sigma is preferentially expressed by myoepithelial cells. Simpson et al. demonstrated that 14-3-3sigma was consistently expressed in the cytoplasmic compartment and occasionally in the nuclei of myoepithelial cells arranged as a continuous layer around normal ducts and lobular units. 14-3-3sigma cytoplasmic subcellular localization was a statistically significant prognostic factor for the whole series of invasive carcinomas, as well as for those positive for ER or PR (65).

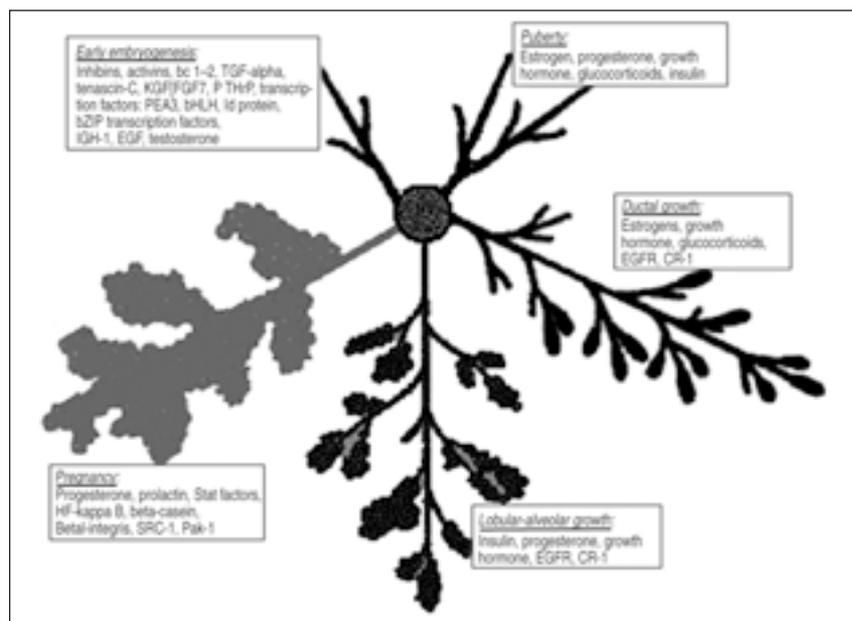


Fig. 1. Schematic representation of mammary gland development

Using an antibody against the P2X7 (apoptotic purinergic receptor), Slater et al. found that normal and mildly hyperplastic epithelium was devoid of the cytolytic P2X7 receptors whereas all epithelial cells in all cases of in situ or invasive lobular or ductal carcinoma labelled intensely. The lobular and ductal in situ cases labelled intracellularly while the invasive epithelial cancer cells showed intense cell surface label indicating an attempt was being made to induce apoptosis (66).

The study of Barsky confirmed the hypothesis that human myoepithelial cells express a distinct tumor-suppressor phenotype. The global gene expression profile (22,000 genes) was examined using Affymetrix Microarray Gene Chips. The myoepithelial cell lines/ xenografts were distinct and very different from nonmyoepithelial breast carcinoma cells and normal breast and breast tumor biopsies. Two hundred and seven specifically selected genes represented a subset of genes that distinguished all the myoepithelial cell lines/xenografts from all the other samples and which themselves exhibited hierarchical clustering (4).

Conclusion

The mammary gland is a complex organ that begins development early in gestation and constantly changes in size, shape and function from the time of puberty to the menopause, affected by varying levels of the female hormone estrogen. All those changes are associated with a variety of hormonal and genetic risk factors and subsequent development of breast cancer. From the embryological point of view there exists no substantial difference between mammary ductal and lobular cells and the morphology of both results from the same developmental process. There is no evidence for a different gene expressing profile in normal ductal and lobular cells. The extensive research currently being performed on the molecular pathology, molecular biology, cancer genomics and proteomics will hopefully contribute to the further elucidation of all genetic and environmental factors involved in the development, differentiation, and involution of the mammary gland. This may give insights into the etiology, pathogenesis, early detection, treatment, and potential prevention of breast cancer.

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References

1. **Anbazhagan, R., Bartek, J., Monaghan, P., et al.:** Growth and development of the human infant breast. *Am. J. Anat.*, 192:407-417, 1991. – 2. **Anderson E.:** The role of oestrogen and progesterone receptors in human mammary development and tumorigenesis. *Breast Cancer Res.*, 4:197-201, 2002. – 3. **Barcellos-Hoff, M.H., Ewan, K.B.:** Transforming growth factor-beta and breast cancer: Mammary gland development. *Breast Cancer Res.*, 2:92-99, 2000. – 4. **Barsky, S.H.:** Myoepithelial mRNA expression profiling reveals a common tumor-suppressor phenotype. *Exp. Mol. Pathol.*, 74:113-122, 2003. – 5. **Chen, H., Sukumar, S.:** Role of homeobox genes in normal mammary gland development and breast tumorigenesis. *J. Mammary Gland Biol. Neoplasia*, 8:159-75, 2003. – 6. **Chepko, G., Smith, G.H.:** Three division-competent, structurally-distinct cell populations contribute to murine mammary epithelial renewal. *Tissue Cell*, 29:239-253, 1997. – 7. **Chughtai, N., Schimchowitsch, S., Lebrun, J.J., et al.:** Prolactin induces SHP-2 association with Stat5, nuclear translocation, and binding to the beta-casein gene promoter in mammary cells. *J. Biol. Chem.*, 277:31107-31114, 2002. – 8. **Clayton, H., Titley, I., Vivanco, M.:** Growth and differentiation of progenitor/stem cells derived from the human mammary gland. *Exp. Cell Res.*, 297:444-60, 2004. – 9. **Desprez, P.Y., Sumida, T., Coppe, J.P.:** Helix-loop-helix proteins in mammary gland development and breast cancer. *J. Mammary Gland Biol. Neoplasia*, 8:225-39, 2003. – 10. **Deugnier, M.A., Faraldo, M.M., Janji, B., et al.:** EGF controls the in vivo developmental potential of a mammary epithelial cell line possessing progenitor properties. *J. Cell Biol.*, 159:453-63, 2002. – 11. **Deugnier, M.A., Moiseyeva, E.P., Thiery, J.P., et al.:** Myoepithelial cell differentiation in the developing mammary gland: progressive acquisition of smooth muscle phenotype. *Dev. Dyn.*, 204:107-117, 1995. – 12. **Dontu, G., Al-Hajj, M., Abdallah, W.M., et al.:** Stem cells in normal breast development and breast cancer. *Cell Prolif.*, 36:59-72, 2003. – 13. **Dunbar, M.E., Wysolmerski, J.J.:** Parathyroid hormone-related protein: a developmental regulatory molecule necessary for mammary gland development. *J. Mammary Gland Biol. Neoplasia*, 4:21-34, 1999. – 14. **Dunbar, M.E., Young, P., Zhang, J.P., et al.:** Stromal cells are critical targets in the regulation of mammary ductal morphogenesis by parathyroid hormone-related protein. *Dev. Biol.*, 203:75-89, 1998. – 15. **Faraldo, M.M., Deugnier, M.A., Tlouzeau, S., et al.:** Perturbation of beta1-integrin function in involuting mammary gland results in premature dedifferentiation of secretory epithelial cells. *Mol. Biol. Cell*, 13:3521-3531, 2002. – 16. **Feuerhake, F., Sigg, W., Hoffer, EA., et al.:** Immunohistochemical analysis of Bcl-2 and Bax expression in relation to cell turnover and epithelial differentiation markers in the non-lactating human mammary gland epithelium. *Cell Tissue Res.*, 299:47-58, 2000. – 17. **Foley, J., Dann, P., Hong, J., et al.:** Parathyroid hormone-related protein maintains mammary epithelial fate and triggers nipple skin differentiation during embryonic breast development. *Development*, 128:513-25, 2001. – 18. **Forster, C., Makela, S., Warri, A., et al.:** Involvement of estrogen receptor beta in terminal differentiation of mammary gland epithelium. *Proc. Natl. Acad. Sci. USA*, 99:15578-83, 2002. – 19. **Geymayer, S., Doppler, W.:** Activation of NF-kappaB p50/p65 is regulated in the developing mammary gland and inhibits STAT5-mediated beta-casein gene expression. *FASEB J.*, 14:1159-70, 2000. – 20. **Gomm, J.J., Smith, J., Ryall, G.K., et al.:** Localization of basic fibroblast growth factor and transforming growth factor beta 1 in the human mammary gland. *Cancer Res.*, 51:4685-92, 1991. – 21. **Grimm, S.L., Rosen, J.M.:** The role of C/EBPbeta in

mammary gland development and breast cancer. *J. Mammary Gland Biol. Neoplasia*, 8:191-204, 2003. – 22. **Ham, A.W., Cormack, D.H.:** The breast. In: *Histology*; 8th ed. Philadelphia: JB Lippincott, 1979:866-874. 23. **Hamilton N.J., Boyd, J.D., Mossman, H.W.:** Human embryology. Cambridge, England, Hetter, 1968. – 24. **Hebbard, L., Steffen, A., Zawadzki, V., et al.:** CD44 expression and regulation during mammary gland development and function. *J. Cell Sci.*, 113:2619-2630, 2000. – 25. **Hennighausen, L., Robinson, G.W.:** Signaling pathways in mammary gland development. *Dev. Cell*, 1:467-475, 2001. – 26. **Hovey, R.C., Trott, J.F., Vonderhaar, B.K.:** Establishing a framework for the functional mammary gland: from endocrinology to morphology. *J. Mammary Gland Biol. Neoplasia*, 7:17-38, 2002. – 27. **Howlett, A.R., Bissell, M.J.:** The influence of tissue microenvironment (stroma and extracellular matrix) on the development and function of mammary epithelium. *Epithelial Cell Biol.*, 2:79-89, 1993. – 28. **Jones, F.E., Welte, T., Fu, X.Y., et al.:** ErbB4 signaling in the mammary gland is required for lobuloalveolar development and Stat5 activation during lactation. *J. Cell Biol.*, 147:77-88, 1999. – 29. **Kazansky, A.V., Raught, B., Lindsey, S.M., Wang, et al.:** Regulation of mammary gland factor/Stat5a during mammary gland development. *Mol. Endocrinol.*, 9:1598-1609, 1995. – 30. **Kordon, E.C., Smith, G.H.:** An entire functional mammary gland may comprise the progeny from a single cell. *Development*, 125:1921-1930, 1998. – 31. **Kumar, R., Wang, R.A.:** Protein kinases in mammary gland development and cancer. *Microsc. Res. Tech.*, 59:49-57, 2002. – 32. **Kurpios, N.A., Sabolic, N.A., Shepherd, T.G., et al.:** Function of PEA3 Ets transcription factors in mammary gland development and oncogenesis. *J. Mammary Gland Biol. Neoplasia*, 8:177-190, 2003. – 33. **LeBrun, D.P., Warnke, R.A., Cleary, M.L.:** Expression of bcl-2 in fetal tissues suggests a role in morphogenesis. *Am. J. Pathol.*, 142:743-753, 1993. – 34. **Lewis, M.T.:** Homeobox genes in mammary gland development and neoplasia. *Breast Cancer Res.*, 2:158-169, 2000. – 35. **Li, M., Liu, X., Robinson, G., Bar-Peled, U., et al.:** Mammary-derived signals activate programmed cell death during the first stage of mammary gland involution. *Proc. Natl. Acad. Sci. USA.* 94:3425-3430, 1997. – 36. **Lin, C.Q., Parrinello, S., Campisi, J., et al.:** Regulation of mammary epithelial cell phenotypes by the helix-loop-helix protein, Id-1. *Endocr. Relat. Cancer*, 6:49-50, 1999. – 37. **Long, W., Wagner, K.U., Lloyd, K.C., et al.:** Impaired differentiation and lactational failure of ErbB4-deficient mammary glands identify ERBB4 as an obligate mediator of STAT5. *Development*, 130:5257-5268, 2003. – 38. **Lu, Q.L., Abel, P., Foster, C.S., et al.:** bcl-2: role in epithelial differentiation and oncogenesis. *Hum. Pathol.*, 27:102-110, 1996. – 39. **Maroulakou, I.G., Bowe, D.B.:** Expression and function of Ets transcription factors in mammalian development: a regulatory network. *Oncogene*, 19:6432-6442, 2000. – 40. **McKiernan, J., Coyne, J., Cahalane, S.:** Histology of breast development in early life. *Arch. Dis. Child.*, 63:136-9, 1988. – 41. **Miyoshi, K., Shillingford, J.M., Smith, G.H., et al.:** Signal transducer and activator of transcription (Stat) 5 controls the proliferation and differentiation of mammary alveolar epithelium. *J. Cell Biol.*, 155:531-542, 2001. – 42. **Monaghan, P., Perusinghe, N.P., Cowen, P., et al.:** Peripubertal human breast development. *Anat Rec.* 226:501-508, 1990. – 43. **Naccarato, A.G., Viacava, P., Vignati, S., et al.:** Bio-morphological events in the development of the human female mammary gland from fetal age to puberty. *Virchows Arch.*, 436:431-438, 2000. – 44. **Nathan, B., Anbazhagan, R., Clarkson, P., et al.:** Expression of BCL-2 in the developing human fetal and infant breast. *Histopathology*, 24:73-76, 1994. – 45. **Neville, M.C., McFadden, T.B., Forsyth, I.:** Hormonal regulation of mammary differentiation and milk secretion. *J. Mammary Gland Biol. Neoplasia*, 7:49-66, 2002. – 46. **Osin, P.P., Anbazhagan, R., Bartkova, J., et al.:** Breast development gives insights into breast disease. *Histopathology*, 33:275-283, 1998. – 47. **Palmieri, C., Roberts-Clark, D., Assadi-Sabet, A., et al.:** Fibroblast growth factor 7, secreted by breast fibroblasts, is an interleukin-1beta-induced paracrine growth factor for human breast cells. *J. Endocrinol.*, 177:65-81, 2003. – 48. **Pechoux, C., Gudjonsson, T., Ronnov-Jessen, L., et al.:** Human mammary luminal epithelial cells contain progenitors to myoepithelial cells. *Dev. Biol.*, 206:88-99, 1999. – 49. **Philp, J.A., Burdon, T.G., Watson, C.J.:** Differential activation of STATs 3 and 5 during mammary gland development. *FEBS Lett.*, 396:77-80, 1996. – 50. **Qi, C., Kashireddy, P., Zhu, Y.T., et al.:** Null mutation of peroxisome proliferator-activated receptor-interacting protein in mammary glands causes defective mammapoiesis. *J. Biol. Chem.*, 279:33696-701, 2004. – 51. **Robinson, G.W., Hennighausen, L.:** Inhibins and activins regulate mammary epithelial cell differentiation through mesenchymal-epithelial interactions. *Development*, 124:2701-8, 1997. – 52. **Robinson, G.W., Karpf, A.B., Kratochwil, K.:** Regulation of mammary gland development by tissue interaction. *J. Mammary Gland Biol. Neoplasia*, 4:9-19, 1999. – 53. **Robinson, G.W., McKnight, R.A., Smith, G.H., et al.:** Mammary epithelial cells undergo secretory differentiation in cycling virgins but require pregnancy for the establishment of terminal differentiation. *Development*, 121:2079-2090, 1995. – 54. **Rosen, P.P.:** *Rosen's Breast Pathology*. Lippincott-Raven Publishers, 1997. – 55. **Rudland, P.S.:** Epithelial stem cells and their possible role in the development of the normal and diseased human breast. *Histol. Histopathol.*, 8:385-404, 1993. – 56. **Rudland, P.S.:** Histochemical organization and cellular composition of ductal buds in developing human breast: evidence of cytochemical intermediates between epithelial and myoepithelial cells. *J. Histochem. Cytochem.*, 39:1471-1484, 1991. – 57. **Russo, J., Ao, X., Grill, C., Russo, I.H.:** Pattern of distribution of cells positive for estrogen receptor alpha and progesterone receptor in relation to proliferating cells in the mammary gland. *Breast Cancer Res. Treat.*, 53:217-227, 1999. – 58. **Russo, J., Hu, Y.F., Silva, I.D., et al.:** Cancer risk related to mammary gland structure and development. *Microsc. Res. Tech.*, 52:204-223, 2001. – 59. **Russo, J., Russo, I.H.:** Development of the human breast. *Maturitas*, 24:2-15, 2004. – 60. **Saji, S., Jensen, E.V., Nilsson, S., et al.:** Estrogen receptors alpha and beta in the rodent mammary gland. *Proc. Natl. Acad. Sci. USA*, 97:337-342, 2000. – 61. **Sakakura, T., Ishihara, A., Yatani, R.:** Tenascin in mammary gland development: from embryogenesis to carcinogenesis. *Cancer Treat. Res.*, 53:383-400, 1991. – 62. **Salomon, D.S., Bianco, C., De Santis, M.:** Cripto: a novel epidermal growth factor (EGF)-related peptide in mammary gland development and neoplasia. *Bioessays*, 21:61-70, 1999. – 63. **Shim, W.S., DiRenzo, J., DeCaprio, J.A., et al.:** Segregation of steroid receptor coactivator-1 from steroid receptors in mammary epithelium. *Proc. Natl. Acad. Sci. USA*, 96:208-213, 1999. – 64. **Shyamala, G., Louie, S.G., Camarillo, I.G., et al.:** The progesterone receptor and its isoforms in mammary development. *Mol. Genet. Metab.*, 68:182-90, 1999. – 65. **Simpson, P.T., Gale, T., Reis-Filho, J.S., et al.:** Distribution and significance of 14-3-3sigma, a novel myoepithelial marker, in normal, benign, and malignant breast tissue. *J. Pathol.*, 202:274-285, 2004. – 66. **Slater, M., Danieletto, S., Pooley, M., et al.:** Differentiation between cancerous and normal hyperplastic lobules in breast lesions. *Breast Cancer Res. Treat.*, 83:1-10, 2004. – 67. **Smith, G.H.:** Experimental mammary epithelial morphogenesis in an in vivo model: evidence for distinct cellular progenitors of the ductal and lobular phenotype. *Breast Cancer Res. Treat.*, 39:21-31, 1996. – 68. **Stingl, J., Eaves, C.J., Kuusk, U., et al.:** Phenotypic and functional characterization in vitro of a multipotent epithelial cell present in the normal adult human breast. *Differentiation*, 63:201-213, 1998. – 69. **Topper, Y.J., Freeman, C.S.:** Multiple hormone interactions in the developmental biology of the mammary gland. *Physiol. Rev.*

60:1049-1106, 1980. – 70. **Van Winter, J.T., Noller, K.L., Zimmerman, D., et al.:** Natural history of premature thelarche in Olmsted County, Minnesota, 1940 to 1984. *J. Pediatr.*, 116:278-280, 1990. – 71. **Visvader, J.E., Lindeman, G.J.:** Transcriptional regulators in mammary gland development and cancer. *Int. J. Biochem. Cell Biol.*, 35:1034-51, 2003. – 72. **Wang, R.A., Vadlamudi, R.K., Bagheri-Yarmand, R., et al.:** Essential functions of p21-activated kinase 1 in morphogenesis and differentiation of mammary glands. *J. Cell Biol.*, 161:583-592, 2003. – 73. **Wiesen, J.F., Young, P., Werb, Z., et al.:** Signaling through the stromal epidermal growth factor receptor is necessary for mammary ductal development. *Development*, 126:335-344, 1999. – 74. **Wiseman, B.S., Werb, Z.:** Stromal effects on mammary gland development and breast cancer. *Science*, 296:1046-1049, 2002. – 75. **Wysolmerski, J.J., McCaughern-Carucci, J.F., Daifotis, A.G., et al.:** Overexpression of parathyroid

hormone-related protein or parathyroid hormone in transgenic mice impairs branching morphogenesis during mammary gland development. *Development*, 121:3539-3547, 1995. – 76. **Yamashita, H., Nevalainen, M.T., Xu, J., et al.:** Role of serine phosphorylation of Stat5a in prolactin-stimulated beta-casein gene expression. *Mol Cell Endocrinol.*, 183:151-163, 2001.

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Jaká je vaše diagnóza?

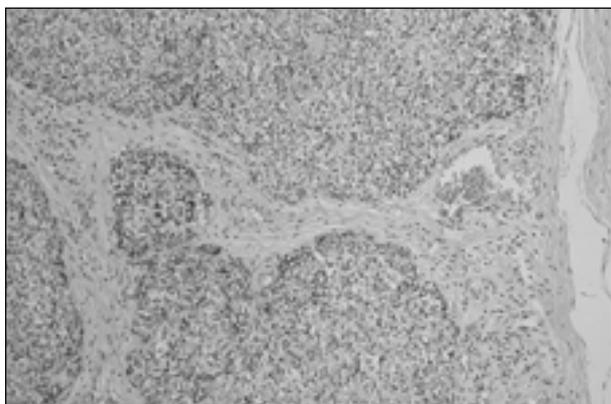
73letá žena s devět měsíců trvajícím nebolestivým zduřením v pravé parotideomaseterické krajině. Při fyzikálním vyšetření byl zjištěn v této lokalizaci kulovitý útvar průměru 40 mm s intaktním kožním krytem. Poruchy funkce příušní slinné žlázy nebyly přítomny, inervace n. VII byla zachována. Ultrasonograficky byl v příušní slinné žláze zastižen hyperechogenní útvar rozměrů 27 x 16 mm a bylo doporučeno jeho odstranění. Při operaci byl zjištěn nádor v povrchovém laloku žlázy, který utlačoval, ale neprorůstal větvení n. VII. Byla provedena pouze exstirpace nádoru, po které následovala aktinoterapie. Devět měsíců po operaci a šest měsíců po poslední kúře aktinoterapie je pacientka bez klinických známek lokální recidivy či generalizace nádorového onemocnění.

K histologickému vyšetření byla zaslána kulovitá částice 35 x 30 x 20 mm s dobře ohraničeným uzlem tužší konzistence průměru 20 mm. Na řezu byl uzel homogenní, šedobílé barvy.

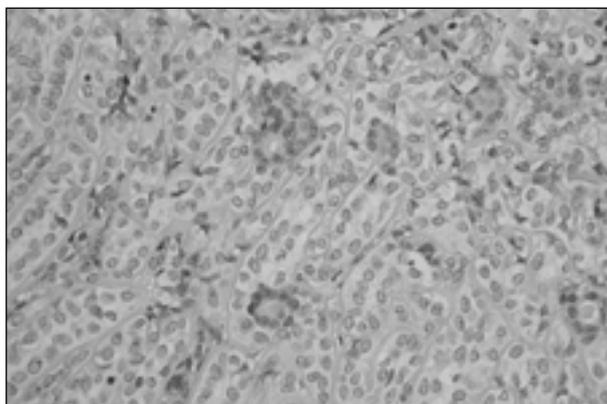
Mikroskopicky byl zastižen vazivově opouzdřený, solidně rostoucí nádor, subkapsulárně lobulárně uspořádaný (obr. 1), který vykazoval v zásadě bifazickou strukturu. Většinu nádorové populace tvořily velké polygonální buňky se světlou cy-

toplazmou, které se nacházely periferně od drobných lumen vystlaných kubickými buňkami s eozinofilní cytoplazmou (obr. 2). V některých lumenech byl přítomen eozinofilní materiál. Stroma nádoru bylo chudé. Nádorové elementy nevykazovaly výraznější jaderné ani cytologické atypie, mitózy byly řídké (2–3 na 10 HPF).

Imunohistochemicky duktální epitelie exprimovaly silně cytokeratiny (CK) a nekonstantně a slabě epitelální membránový antigen (EMA) a S-100 protein. Světlobuněčná komponenta nádoru naopak silně exprimovala hladkosvalový aktin (SMA), protein p63 a S-100 protein; vimentin byl exprimován pouze fokálně. Stroma nádoru bylo silně PAS pozitivní, stejně jako materiál v drobných lumenech. Světlobuněčná komponenta nádoru byla PAS negativní. Stroma nádoru, stejně jako materiál v drobných lumenech, vykazovalo též pozitivitu při stříbření dle Jonese. Výsledek barvení na hlen byl negativní, stejně jako imunohistochemický průkaz GFAP. Proliferační antigen Ki-67 byl exprimován v nečetných buňkách, a to výhradně v buňkách se světlou cytoplazmou (do 5 %), převážně v periferních, resp. subkapsulárních partiích nádoru.



Obr. 1. Solidně rostoucí, ložiskově lobulárně uspořádaný nádor je obdán vazivovým pouzdrem, z něhož vycházejí do nádorové tkáně neúplná septa (HE, 100krát)



Obr. 2. Bifazické uspořádání nádoru (HE, 400krát)

Odpověď na str. 118